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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OF

### FORM PTO-1390 (Modified) TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1 5

INTERNATIONAL APPLICATION NO PCT/GB98/02550

INTERNATIONAL FILING DATE 21 August 1998 - PRIORITY DATE CI 22 August 1997

LABELLING COMPOSITION AND METHOD >

| APPLICANT(S) FOR DO/EO/US | 3 |
|---------------------------|---|
| HOPKINS, Alison           |   |

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

- This is a FIRST submission of items concerning a filing under 35 U.S.C. 371
- This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.
- This is an express request to begin national examination procedures (35 U.S C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
- A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. X 4
- A copy of the International Application as filed (35 U S.C. 371 (c) (2))
  - is transmitted herewith (required only if not transmitted by the International Bureau).
    - has been transmitted by the International Bureau
  - is not required, as the application was filed in the United States Receiving Office (RO/US).
  - A translation of the International Application into English (35 U.S.C. 371(c)(2)).
- A copy of the International Search Report (PCT/ISA/210).
  - Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
    - are transmitted herewith (required only if not transmitted by the International Bureau)
    - have been transmitted by the International Bureau.
    - have not been made; however, the time limit for making such amendments has NOT expired.
    - A have not been made and will not be made.
- ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
- An oath or declaration of the inventor(s) (35 U.S C. 371 (c)(4)) 10
- A copy of the International Preliminary Examination Report (PCT/IPEA/409).
- A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 12 (35 U.S.C. 371 (c)(5)).

### Items 13 to 20 below concern document(s) or information included:

- An Information Disclosure Statement under 37 CFR I 97 and 1 98 13.
- An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3 28 and 3.31 is included. 14
- 15 A FIRST preliminary amendment
- A SE 16
- 17 A sub
- 18 A cha
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- 20 Other

| COND or SUBSEQUENT preliminary amendment.       |
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| <ol> <li>The following fees are submitted</li> </ol>  | : -                            |  |                  |                     |                   | CALCULATIONS              | PTO USE ONLY |
| BASIC NATIONAL FEE ( 37 CFR 1.492 (   | a) (1) -                       | (5)):  |                  |                     | - 1               |                           |              |
| ☐ Neither international preliminary examinternational search fee (37 CFR 1.44 and International Search Report not present the control of the | mination<br>5(a)(2)<br>repared | n fee (37 CFR 1 482) nor<br>paid to USPTO<br>by the EPO or JPO |                  | s                   | 970.00            |                           |              |
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| ☐ International preluminary examination<br>but all claims did not satisfy provision   | ns of PC                       | T Article 33(1)-(4)  |                  | . 8                 | 670.00            |                           |              |
| <ul> <li>International preliminary examination<br/>and all claims satisfied provisions of</li> </ul>  | PCT Ar                         | ticle 33(1)-(4)  |                  | OT1317              | \$96.00           |                           |              |
|   |                                | ATE BASIC FEE  |                  |                     |                   | \$840,00                  |              |
| Surcharge of \$130.00 for furnishing the oath<br>months from the earliest claimed priority da   | te (37 C                       | FR 1.492 (e)).   | ☐ 20             |                     | 30                | \$130.00                  |              |
| CLAIMS NUMBER FILE  |                                | NUMBER EXTRA   | $\dashv$         | R.A                 | 8.00              | \$0.00                    |              |
| Total claims 6 - 20<br>Independent claims 1 -   |                                | 0  | -                |                     | 8.00              | \$0.00                    |              |
| Independent claims 1 - Multiple Dependent Claims (check if applie   |                                | L  | -                | , , , ,             |                   | \$0.00                    |              |
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| Reduction of 1/2 for filing by small entity, it<br>must also be filed (Note 37 CFR 1 9, 1.27, 1   | f applica                      | ble. Verified Small Enti                                       |                  |                     |                   | \$0.00                    |              |
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| Fee for recording the enclosed assignment (3 accompanied by an appropriate cover sheet (  | 37 CFR<br>37 CFR               | 1 21(h)). The assignmen<br>3.28, 3.31) (check if ap            | t must           | be<br>ole).         |                   | \$0.00                    |              |
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## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

| Applicant:                            | ) | "EXPRESS MAIL" mailing label                 |
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|                                       | ) | No. EM362733790US                            |
| Alison Hopkins                        | ) |  |
|                                       | ) | Date of Deposit: February 7, 2000            |
| U.S. National Phase (35 USC 371)      | ) |  |
| Application Based on PCT/GB98/02550   | ) | I hereby certify that this paper (or fee) is |
| filed 21 August 1998                  | ) | being deposited with the United States       |
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|                                       | ) | 37 C.F.R. §1.10 on the date indicated        |
| Filed: Herewith                       | ) | above and is addressed to the Assistant      |
|                                       | ) | Commissioner for Patents, Washington,        |
| For: Labelling Composition and Method | ) | D.C. 20231.                                  |
|                                       | ) | , ,//  |
| Group Art Unit: To Be Determined      | ) | 0/ 1/  |
|                                       | ) | Kehaul Limm                                  |
| Examiner: To Be Determined            | ) | Richard Zimmermann                           |
|                                       |   |  |

# PRELIMINARY AMENDMENT ACCOMPANYING FILING OF NATIONAL STAGE APPLICATION UNDER 35 U.S.C. 371

Box PCT Assistant Commissioner for Patents Washington, D.C. 20231

Sir:

### **AMENDMENT**

### In the Claims:

Please amend claims 3, 4 and 5 as follows:

- --3. [AMENDED] A labelling composition as claimed in claim 1 [or claim
- 2], wherein the random mixture is of 6-mer oligonucleotides .--

--4. [AMENDED] A labelling composition as claimed in <u>claim 1</u> [any one of claims 1 to 3], wherein the composition is present in a freeze-dried state.--

--5. [AMENDED] A method of making labelled probes for a nucleic acid template, which method comprises incubating the nucleic acid template under chain extension conditions with the labelling composition of claim 1 [any one of claims 1 to 4].--

### REMARKS

The foregoing amendments are made to eliminate various multiple dependencies and to place the claims in condition for allowance. An early notice of allowance is hereby solicited. Should the Examiner wish to discuss any matter of form or substance, he or she is invited to contact the undersigned attorney at the number listed below.

Respectfully submitted,

MARSHALL, O'TOOLE, GERSTEIN, MURRAY & BORUN

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Chicago, Illinois February 7, 2000

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### LABELLING COMPOSITION AND METHOD

This invention concerns compositions comprising random mixtures of oligonucleotides and their use for labelling nucleic acids by a random prime method.

Feinberg and Vogelstein (1, 2) introduced the use of random sequence hexanucleotides to prime DNA synthesis on denatured template DNA at numerous sites along its length. The primer-template complex is a substrate for the "Klenow" fragment of DNA polymerase I. By replacing a non-radioactive nucleotide with the radiolabelled equivalent in the reaction mixture, newly synthesised DNA is made radioactive.

Very small amounts of input DNA can be labelled, enabling very high specific activity probes to be produced with relatively small quantities of added nucleotides. These radioactive labelled fragments can then be used as sensitive hybridisation probes for a wide range of filter based applications (3-6).

There are several labelling kits that are commercially available for the labelling of DNA by the random prime method. These include the Multiprime, Megaprime, Rediprime and Fluorescein Gene Images kits available from Amersham International plc. Ready-To-Go kits are available from Pharmacia and High Prime kits are available from Boehringer.

The Multiprime kit was introduced in the 1980s. It provides different tubes containing the different solutions that enable the user to make up labelling mixtures. One such tube contains a random mixture of 6-mer oligonucleotides, another the polymerase enzyme, and another the supply of nucleotides in the reaction buffer. All these separate solutions are stored frozen at -20°C. The purchaser thaws the different solutions, and adds precise quantities of each to the sample of denatured DNA that is

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to be labelled, including a labelled nucleotide. This reaction is then usually incubated at 37°C at which temperature, oligonucleotide annealing and chain extension can occur. However, the reaction may also be incubated at lower temperatures such as an ambient room temperature of about 20°C.

The Megaprime kit was introduced commercially in the early 1990s. It is similar to the Multiprime kit, except that 9-mer oligonucleotides are used in place of 6-mers. The Megaprime kit has an advantage over the Multiprime kit, in that 9-mer oligonucleotides anneal more strongly (than do 6-mers) to a DNA target and form a hybrid having a higher melting temperature. Thus 9-mers achieve better and more rapid priming of a target then do 6-mers.

The Rediprime kit was introduced commercially in 1994. It comprises a mixture of 9-mer oligonucleotides with a polymerase enzyme and a supply of nucleotides. The mixture is supplied in a freeze-dried state. The freeze-dried mixture also contains a dye for easy visualisation. Dried kits for performing nucleic acid manipulation experiments were described by Ortlepp and McKay in EP 298 669 entitled "Performing nucleic acid reactions". The user reconstitutes the mixture by adding liquid containing the DNA template that is to be labelled, and then liquid containing the labelled nucleotide.

The Ready-To-Go kit was introduced during the 1990s. It is based on a random prime solution containing a random mixture of 9-mer or longer oligonucleotides, which solution is dried by a technique described in EP 383 569. A dye is not present. Like the Rediprime kit, the Ready-To-Go kit can be stored at +4°C or at ambient temperature. Promotional literature emphasises the speed of labelling, which results from the use of 9-mer oligonucleotides.

The High Prime kit is a wet kit containing a random mixture of oligonucleotides. The kit literature does not indicate what length of random oligonucleotides are used, but in the related document EP 649 909 A2, the

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use of 6-mer, 9-mer, 12-mer and 15-mer is disclosed. No preferred length of random oligonucleotide is given. The solution is stablised by the use of glycerol and can be stored at between about -20°C and +4°C.

It can be seen that there has been a trend in commercial kits towards the use of longer oligonucleotides, particularly 9-mers or even longer. Going against this trend, it has been determined by Suganuma, A and Gupta, K C (7) that the use of long random primers, especially 9-mers or longer, reduces the priming efficiency of the random primer reaction. These authors worked on solutions which were used without being dried at any stage. The conclusions of these authors conflict with the findings of the present inventors; which findings are to the effect that, when experiments are done with solutions which are not dried, 9-mers provide more rapid and efficient labelling than do 6-mers, and do not give rise to any problem resulting from self-annealing or self-priming. To the best of applicants' knowledge, the conclusions reported by the authors of (7) have not caused the suppliers of random prime kits to use shorter oligonucleotides.

The present invention is based on the discovery that selfannealing occurs when random 9-mers are used in dried predispensed
labelling kits, and that this limits their stability and shelf life. The selfannealing occurs during dispensing and storage when the random 9-mers
anneal together to form primer-dimers or primer concatemers. These
primer complexes become labelled during the normal labelling reaction,
which concomitantly reduces the amount of label that is incorporated into
copies of the template that are being synthesised during the reaction.
Shorter oligonucleotides are not subject to this problem. The problem is
specific to 9-mers (and longer oligonucleotides) used in dried kits.

The invention provides a labelling composition comprising a random mixture of oligonucleotides which are 6-mers to 8-mers, said composition present in a dry state. Preferably the composition also contains at least one of: a polymerase enzyme; a supply of nucleotides for

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chain extension; a labelled nucleotide; a dve; a stabiliser; and a buffer.

As the experimental data below shows, 5-mer oligonucleotides are too short to be useful in dried kits. As the length of the oligonucleotides increases from 6-mers to 9-mers, there is a concomitant increase in the aforementioned self-priming problem. On the other hand, longer oligonucleotides anneal more rapidly and strongly to templates than do shorter ones. Taking into account both these factors, applicants believe that 6-mer oligonucleotides are more preferable than 7-mers which in turn are more preferable than 8-mers.

The random mixture of oligonucleotides is present in a dry state. Various drying techniques are possible, including that described in EP 383 569, and also freeze-drying or Ivophilisation which is preferred.

It is possible to use any DNA polymerase enzyme in the labelling reaction, for example Klenow, exonuclease free klenow, DNA polymerase I, T7 DNA polymerase, Sequenase<sup>™</sup>, Thermosequenase<sup>™</sup>, so long as the reaction buffer conditions are suitable for the specific enzyme being used.

All four of the nucleotides are preferably present in the composition, whether labelled or unlabelled, and the relative molar concentrations may be adjusted to improve the efficiency of labelling. Also when a labelled nucleotide is present, the equivalent unlabelled nucleotide may also be present to improve the efficiency of labelling, or to control the specific activity of the DNA that is being produced from the labelling reaction.

These compositions enable a DNA template to be used to produce copies which are labelled radioactively, for example, by using either phosphate labelled with P-32 or S-35, or by using H-3 or C-14 base labelled nucleotides. Alternatively non-radioactive labels may be used, for example, fluorescein, biotin, digoxigenin, rhodamine and cyanine dyes, may be incorporated when, for example, covalently linked to the base

moiety of the nucleotide.

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Any stabiliser may be present to protect the activity of the enzyme, for example, trehalose, sucrose, BSA, gelatin. A dye may also be present to allow the dried pellet to be visualised, before use, and to assist in determining that mixing is thorough.

The invention also includes a method of making labelled probes for a nucleic acid template, which method comprises incubating the nucleic acid template under chain extension conditions with the labelling composition as herein described. Preferably the template is DNA. The inventor has found that random 6-mers can give fast labelling kinetics (10 minutes labelling time) by being present at high concentration in the reaction mixture. A preferred concentration is 2-10 O.D./ml in the final reaction with about 5 O.D./ml being most preferable. A probe labelled in this manner is suitable for use in a Southern hybridisation.

All the results shown in the examples show labelling with dCTP-<sup>32</sup>P, but this is only as a means to show, and quantitate the amount of self-priming that occurred in each reaction. The reactions are able to label DNA with other labels, both radioactive and non-radioactive, as indicated elsewhere in this specification.

### 20 References

- Feinberg, A P and Vogelstein, B, Anal. Biochem., 132: 6-13 (1983).
- 2. Feinberg, A P and Vogelstein, B, Addendum Anal. Biochem., 137: 266-267 (1984).
- 25 3. Southern, E M, J. Mol. Biol., 98: 503-517 (1975).
  - Thomas, P S, Proc. Nat. Acad. Sci., USA, 77: 5201-5205
     (1980).
  - Meinkoth, J and Wahl, G, Anal. Biochem, 138: 267-284
     (1984).
- Grunstein, M and Hogness, D S, Proc. Natl. Acad. Sci, USA,
   3961-3965 (1975).

7. Sugunuma, A and Gupta, K C, Analytical Biochemistry, 224: 605-608 (1995).

# Example 1. Manufacture of lyophilised reactions with different random primer lengths:

All primers were diluted to 50 O.D./ml in water. The number of enzyme units was the same in each reaction (7 units).

The amount of each component solution is as follows for a 6 ml scale.

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|   | 5 mer<br>reaction mix | 6 mer<br>reaction mix | 7 mer<br>reaction mix | 8 mer<br>reaction mix | 9 mer<br>reaction mix |
|---|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|
| Nucleotide buffer                       | 1.998 ml              |
| Exo-free Klenow<br>(12 μl) 100 units/μl | 1200 units            |
| Dilution Buffer                         | 28 μΙ                 | 28 μΙ                 | 28 µl                 | 28 µl                 | 28 µl                 |
| 5 mer primer                            | 1.0 ml                |                       |                       |                       |                       |
| 6 mer primer                            |                       | 1.0 ml                |                       |                       |                       |
| 7 mer primer                            |                       |                       | 1.0 ml                |                       |                       |
| 8 mer primer                            |                       |                       |                       | 1.0 ml                |                       |
| 9 mer primer                            |                       |                       |                       |                       | 1.0 ml                |
| 20% Trehalose                           | 1.5 ml                |
| 0.2 mg/ml Xylene<br>Cyanol              | 0.198 ml              | 0.198 mi              | 0.198 ml              | 0.198 ml              | 0.198 ml              |
| PF Water                                | 1.264 ml              |
| Total Volume                            | 6 ml                  |

Each reaction mix was dispensed into tubes in 35  $\mu$ l aliquots, and were freeze dried.

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### Methods:

- Nucleotide buffer: Labelling buffer from Nick Translation kit 1. (N5000/N5500 Amersham International plc).
- 2. Dilution buffer: Storage buffer for enzyme dilution.
- 3 Labelling Method: Tubes of DNA for labelling were made up as follows:

5 μl λ HindIII DNA at 5 ng/μl in TE buffer.

40 ul water.

Placed all tubes in a boiling water bath (95 to 100°C) for 10 5 minutes.

placed all tubes on ice for 5 minutes, centrifuged briefly. then added the denatured DNA solutions to the respective dried reaction tube samples

added 5 µl Redivue™ dCTP (α32P) (Product Code AA0005:

Amersham International plc) (50 µl total reaction volume).

Incubated all reactions for 10 minutes at 37°C.

Spotted 2 µl samples out onto PEI-cellulose tlc plates,

Ran plates in 1.25 M KH<sub>2</sub>PO<sub>4</sub> pH 3.4.

Analysed plates on plate scanner, to measure the %incorporation, %self-priming and %dCTP present at the end of each reaction.

The %self-priming is defined as the % of the total radioactive counts that are situated between the incorporated counts and the counts due to the unincorporated dCTP-32P.

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## λ HindIII DNA Labelling with dCTP-32P (Week 1 Test)

|       |                |             | Tube-1           |           | Tube-2      |                  |           |  |
|-------|----------------|-------------|------------------|-----------|-------------|------------------|-----------|--|
| Tube  | Primer<br>Type | %<br>Incorp | % Self-<br>Prime | %<br>dCTP | %<br>Incorp | % Self-<br>Prime | %<br>dCTP |  |
| 1, 2  | 5 mers         | 62.7        | 7.9              | 23.0      | 54.8        | 7.7              | 30.9      |  |
| 3, 4  | 6 mers         | 79.9        | 11.2             | 2.7       | 82.1        | 10.8             | 2.2       |  |
| 5, 6  | 7 mers         | 73.5        | 17.5             | 2.8       | 74.3        | 15.1             | 3.7       |  |
| 7, 8  | 8 mers         | 68.3        | 19.1             | 3.2       | 65.6        | 20.4             | 3.6       |  |
| 9, 10 | 9 mers         | 64.9        | 23.7             | 3.1       | 61.5        | 27.2             | 3.1       |  |

The column headed "% Incorp" shows the percentage of dCTP-<sup>32</sup>P incorporated as a chain extension product of a primer-λHind III DNA hybrid. The column headed "% Self-Prime" shows the percentage of dCTP-<sup>32</sup>P incorporated in a complex involving only primers. The column headed "% dCTP" shows the percent of unincorporated dCTP-<sup>32</sup>P. The % dCTP figures were unacceptably high when 5-mer oligonucleotides were used, but were acceptable for 6-mers to 9-mers. Within this range, the % Incorp figures decrease as the oligonucleotide length increases from 6 to 9.

Example 2. Long term stability comparison of dried reactions,

nonamers compared with hexamers, 3.5 units of Exo-free Klenow per reaction:

The samples were made up as shown in Example 1, but 6  $\mu$ l of Exo-free Klenow was used

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# DNA Labelling with dCTP-<sup>32</sup>P, results are the averages of the three reactions

|      |          | Nonamers         |        | Hexamers |                  |        |
|------|----------|------------------|--------|----------|------------------|--------|
| Week | % Incorp | % Self-<br>Prime | % dCTP | % Incorp | % Self-<br>Prime | % dCTP |
| 3    | 61.9     | 17.5             | 6.2    | 69.6     | 9.7              | 6.3    |
| 6    | 71.4     | 18.0             | 4.3    | 80.8     | 8.4              | 4.7    |
| 10   | 65.8     | 20.2             | 6.4    | 75.0     | 11.9             | 6.9    |
| 16   | 66.5     | 16.5             | 8.0    | 73.4     | 11.1             | 6.5    |
| 21   | 78.3     | 10.8             | 3.0    | 84.3     | 5.8              | 2.4    |
| 25   | 42.7     | 11.6             | 40.4   | 55.1     | 5.4              | 35.1   |

As these figures show, the % incorporation of dCTP-<sup>32</sup>P when using 9-mers was initially lower than when using 6-mers and remained lower on storage of the compositions for up to 25 weeks.

### Example 3:

Using dried reactions as shown in Example 1, the primer was replaced with water for the reaction drying, and was added later as a separate solution, when the reactions were being used. All reactions were incubated for 10 minutes, and then sampled to measure the % incorporation.

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| Primer Concentration | % Incorporation          | % Incorporation   |
|----------------------|--------------------------|-------------------|
| in reaction          | (hexamer primers)        | (nonamer primers) |
| O.D./ml              | Average of two reactions | one reaction      |
| 6.0                  | 78.3                     |                   |
| 5.0                  | 83.2                     | 81.0              |
| 4.0                  | 67.7                     | 65.6              |
| 2.0                  | 51.5                     | 67.0              |
| 1.0                  | 45.1                     | 60.2              |

It can be seen from these results that the same primer concentration (O.D./ml) is required to achieve the same reaction kinetics, i.e. the same % incorporation in 10 minutes with different random primer lengths. This shows that the molar concentration needs to increase as the primer length is reduced.

Although the above results were obtained using wet reagents, the conclusion would apply also when dry primers are used.

### Example 4:

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Densitometer results of Southern hybridisations

25ng labelling reactions were carried out using the Megaprime Labelling Kit RPN 1606 (Amersham International plc) or using labelled probes from dried nonamer or hexamer labelling reactions made as described above in other examples. Southern blots were hybridised for 2 hours at 65°C with the labelled probe under standard conditions and then washed in 2 x SSC, 0.1% SDS, 20 minutes at room temperature, followed by two washes in 0.5 x SSC, 0.1% SDS, for 5 minutes 65°C. The dried blots were detected on X-ray film with 2 intensifying screens and place into a -70°C freezer, for 16 hours. After the film was developed using a film processor it was scanned using a densitometer, then the results were analysed using ImageQuant software.

| Kit   | Time of test<br>after<br>manufacture | Target | %band intensity of<br>Southern hybridisation<br>of Megaprime control |
|-------|--------------------------------------|--------|--|
| 9mers | 1 week                               | 0.25pg | 42.23  |
| 9mers | 1 week                               | 0.5pg  | 40.12  |
| 9mers | 1 week                               | 1.0pg  | 38.93  |
| 6mers | 1 week                               | 0.25pg | 97.09  |
| 6mers | 1 week                               | 0.5pg  | 95.02  |
| 6mers | 1 week                               | 1.0pg  | 94.33  |
| 6mers | 37 weeks                             | 0.25pg | 74.58  |
| 6mers | 37 weeks                             | 0.5pg  | 80.91  |
| 6mers | 37 weeks                             | 1.0pg  | 81.17  |

### Conclusions:

The hexamers used in a dried labelling reaction generate labelled probes which gave a much stronger band intensity than when nonamers are used, not only when tested initially after 1 week, but even after an extended period of storage (37 weeks at room temperature).

#### CLAIMS

- A labelling composition comprising a random mixture of oligonucleotides which are 6-mers to 8-mers, said composition present in a dry state.
  - A labelling composition as claimed in claim 1, wherein the composition also contains at least one of: a polymerase enzyme; a supply of nucleotides for chain extension; a labelled nucleotide; a dye; a stabiliser; and a buffer.
  - 3. A labelling composition as claimed in claim 1 or claim 2, wherein the random mixture is of 6-mer oligonucleotides.
  - 4. A labelling composition as claimed in any one of claims 1 to
- 15 3, wherein the composition is present in a freeze-dried state.
  - A method of making labelled probes for a nucleic acid template, which method comprises incubating the nucleic acid template under chain extension conditions with the labelling composition of any one of claims 1 to 4.
- 20 6. A method as claimed in claim 5, wherein the random mixture of oligonucleotides is present at a concentration of 2-10 O.D./ml.

### DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY

| next to my name; I believe that I ar     | n the original, first and sole inventor   | (if only one name is listed below)   | or an orig   | ginal, first |
|--|---|--------------------------------------|--------------|--------------|
| and joint inventor (if plural names a    | are listed below) of the subject matter   | which is claimed and for which a     | patent is    | sought on    |
| the invention entitled ". LABELLI        | NG COMPOSITION AND METHOD   | <u> </u>                             |              |              |
|  | ," the  | specification of which (check one    | ): [] i      | s attached   |
| hereto: was filed on                     | as Application Seria  | •                                    | -            | nended on    |
|  | was filed as PCT International A  |                                      |              |              |
|  | 9 on (if ap   | • •                                  |              |              |
|  | e-identified specification, including the   |                                      |              |              |
|  | disclose to the Patent and Trademark  |                                      |              |              |
| - · · · · ·                              |   | Office all information known to h    |              | naterial to  |
| patentability as defined in 37 C.F.R     | . §1.30.  |                                      |              |              |
| I hereby claim foreign pri               | ority benefits under 35 U.S.C. §119   | of any foreign application(s) for    | patent or    | inventor's   |
|  | onal application(s) designating at least  |                                      |              |              |
| •  | ified below any foreign application(  |                                      |              |              |
|  | ing at least one country other than th  |                                      |              |              |
| **                                       | efore that of the application(s) of which   |                                      | -,           |              |
| subject matter maxing a ming and o       |   |                                      | Priorit      | y Claimed    |
|  |   |                                      |              |              |
| 9717972.5<br>(Application Serial Number) | GB (Country)  | 22-08-1997<br>(Day/Month/Year Filed) | Yes          | No           |
|  |   |                                      |              |              |
|  |   |                                      |              |              |
|  |   |                                      |              |              |
| (Application Serial Number)              | (Country)   | (Day/Month/Year Filed)               | Yes          | No.          |
| ( P                                      | ,,,   | , ,                                  |              |              |
|  |   |                                      |              |              |
| I hereby claim the benefit               | under 35 U.S.C. §119(e) of any Unite  | d States provisional application(s)  | isted belo   | w:           |
|  |   |                                      |              |              |
| (Application Serial Number)              |   | (Day/Month/Year Filed)               |              |              |
|  |   |                                      |              |              |
|  |   |                                      |              |              |
| (Application Serial Number)              |   | (Day/Month/Year Filed)               |              |              |
| V-7E                                     |   | , .                                  |              |              |
| I hereby claim the benefit               | under 35 U.S.C. §120 of any United  | States application(s) or PCT interna | ational app  | olication(s) |
|  | nerica listed below and, insofar as the<br>ation(s) in the manner provided by the |                                      |              |              |
| duty to disclose to the Office all in    | nformation known to me to be materia  | al to patentability as defined in 37 | C.F.R. §     | 1.56 which   |
| occurred between the filing date of      | the prior application(s) and the nation   | al or PCT international filing date  | of this app  | olication:   |
| (Application Serial Number)              | (Day/Month/Year Filed)  | (Status-Patentec                     | I. Pending o | r Abandoned  |
| ·-pp                                     | (Supplied Lott A Body   | (manufic a finance)                  | .,           |              |
|  |   |                                      |              |              |
| (Application Serial Number)              | (Day/Month/Year Filed)  | (Status-Patentee                     |              |              |

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: I hereby appoint as my attorneys, with full powers of substitution and revocation, to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

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| State or Country             |  |
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### APPENDIX

- 1. A labelling composition comprising a random mixture of oligonucleotides which are 6-mers to 8-mers, said composition present in a dry state.
- 2. A labelling composition as claimed in claim 1, wherein the composition also contains at least one of: a polymerase enzyme; a supply of nucleotides for chain extension; a labelled nucleotide; a dye; a stabiliser; and a buffer.
- 3. A labelling composition as claimed in claim 1, wherein the random mixture is of 6-mer oligonucleotides.
- $\mbox{4.} \qquad \mbox{A labelling composition as claimed in claim 1, wherein the composition is present in a freeze-dried state.}$
- 5. A method of making labelled probes for a nucleic acid template, which method comprises incubating the nucleic acid template under chain extension conditions with the labelling composition of claim 1.
- 6. A method as claimed in claim 5, wherein the random mixture of oligonucleotides is present at a concentration of 2-10 O.D./ml.